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(21) International Application Number: PCT/US99/09122 (22) International Filing Date: 27 April 1999 (27.04.99) (30) Priority Data: 09/069,628 29 April 1998 (29.04.98) US (71) Applicant: THE UAB RESEARCH FOUNDATION [US/US]; Suite 1120G/AB, 701 South 20th Street, Birmingham, AL 35294-0111 (US). (72) Inventor: KEARNEY, John, F.; 1430 33rd Street South, Birmingham, AL 35205 (US). (74) Agent: ADLER, Benjamin, A.; McGregor & Adler, LLP, 8011 Candle Lane, Houston, TX 77071 (US).		(81) Designated States: AU, CA, JP, European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
(54) Title: MONOCLONAL ANTIBODIES SPECIFIC FOR ANTHRAX SPORES AND PEPTIDES DERIVED FROM THE ANTIBODIES THEREOF (57) Abstract <p>The present invention provides monoclonal antibodies which are highly specific for <i>Bacillus</i> spores. Also provided are peptides derived from those monoclonal antibodies. Both the antibodies and peptides are highly specific and can discriminate between spores of potentially lethal organisms such as <i>Bacillus anthracis</i> and other harmless but closely related bacilli and provide a very powerful tool in the construction of detection instruments as counter measures.</p> <p>16, + 18 102</p> <p>11-20 123456789101112131415161718192021222324252627282930313233343536373839404142434445464748495051525354555657585960616263646566676869707172737475767778798081828384858687888990919293949596979899100</p>		

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MONOCLONAL ANTIBODIES SPECIFIC FOR ANTHRAX SPORES AND PEPTIDES DERIVED FROM THE ANTIBODIES THEREOF

5

10

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates generally to the fields of immunology and microbiology. More specifically, the present invention relates to monoclonal antibodies specific for anthrax spores and peptides derived from the antibodies.

Description of the Related Art

During the evolution of the immune system there is evidence that the repertoire of germline V genes that has been retained in the genome has been subject to selective processes by

environmental influences which may include self as well as commensal and non-commensal microorganisms. Structural and functional analysis of immunoglobulin and T cell receptors have delineated regions of these molecules which are germline encoded and have the ability to bind to certain bacterial components through exposed parts of the molecules which do not need somatic diversification for expression of the ability to bind to these structures. Some of these included protein A binding to framework three (FR3) region of V_H genes, staphylococcal enterotoxin binding to T cell receptor, etc.

The prior art is deficient in the lack of monoclonal antibodies which are highly specific and can discriminate between spores of the *Bacillus* family including the strategically important *B. anthracis*. Further, the prior art is deficient in the lack of peptides derived from the monoclonal antibodies highly specific for *Bacillus* spores. The present invention fulfills this longstanding need and desire in the art.

SUMMARY OF THE INVENTION

The present invention is directed to monoclonal antibodies which are highly specific for anthrax spores and peptides derived

from the antibodies. The present invention demonstrated that the humoral immune response to spores of *Bacillus* show a remarkable conservation of V_H gene usage which is distinct for each spore analyzed. The results imply evolutionary conservation of V_H genes
5 due to their ability to bind spores. Furthermore, of highly practical importance, these antibodies can discriminate between spores of potentially lethal organisms such as *B. anthracis* and other harmless but closely related bacilli and provide a very powerful tool in the construction of detection instruments as counter measures in
10 biological warfare.

In one embodiment of the present invention, there is provided a monoclonal antibody specific for *Bacillus* spores. Preferably, *Bacillus* is selected from the group consisting of *Bacillus anthracis*, *Bacillus thuringiensis*, *Bacillus subtilis* and other bacilli
15 related to *Bacillus anthracis*. Preferably, the antibody is IgG.

In another embodiment of the present invention, there is provided a peptide derived from the monoclonal antibody highly specific for *Bacillus* spores.

In yet another embodiment of the present invention,
20 there is provided a method of preparing the monoclonal antibody highly specific for *Bacillus* spores by immunizing and fusing local lymph nodes of an animal.

In still yet another embodiment of the present invention, there is provided a method of detecting *Bacillus* spores in a field sample using a monoclonal antibody highly specific for the *Bacillus* spores by contacting the sample with a monoclonal antibody disclosed
5 herein.

In still yet another embodiment of the present invention, there is provided a method of detecting *Bacillus* spores in a field sample using a peptide derived from a monoclonal antibody highly specific for the *Bacillus* spores by contacting the sample with a
10 peptide derived from a monoclonal antibody highly specific for *Bacillus* spores.

Other and further aspects, features, and advantages of the present invention will be apparent from the following description of the presently preferred embodiments of the invention given for the
15 purpose of disclosure.

BRIEF DESCRIPTION OF THE DRAWINGS

So that the matter in which the above-recited features.
20 advantages and objects of the invention, as well as others which will become clear, are attained and can be understood in detail, more particular descriptions of the invention briefly summarized above

may be had by reference to certain embodiments thereof which are illustrated in the appended drawings. These drawings form a part of the specification. It is to be noted, however, that the appended drawings illustrate preferred embodiments of the invention and
5 therefore are not to be considered limiting in their scope.

Figure 1 shows antibodies with different reactivities for germinated and ungerminated spores among different clones.

Figure 2 shows that anti-spore monoclonal antibodies do not react with vegetative bacteria.

10 **Figure 3** shows that most antibodies react specifically with spores of *Bacillus subtilis*.

Figure 4 shows that a commonly used member V_H7183.6 heavy chain gene of the V_H7183 (MOPC21) family in all hybridomas reactive with *Bacillus subtilis* spores appears to be the most unique
15 member of this family in the framework three (FR3) region. (Hybridomas g07 to f10 are labeled SEQ ID NO: 7 to SEQ ID NO: 17, respectively)

Figure 5 shows that antibody-derived peptides specifically bind *Bacillus subtilis* spores.

20 **Figure 6** shows that anti-*Bacillus anthracis* antibody specifically bind *Bacillus anthracis* spores.

Figure 7 shows that the V_H gene sequences among

monoclonal antibodies to *Bacillus anthracis*. (SEQ ID NO: 18 to SEQ ID NO: 31)

Figure 8 shows the discriminatory ability of the antibodies *in vitro*. *Bacillus anthracis*, *Bacillus subtilis* and *Bacillus*
5 *thuringiensis* spores were mixed and stained on a slide with fluorescent antibodies labeled with blue, green and red antibodies, respectively.

Figure 9 shows the discriminatory ability of the antibodies *in vivo*. A section of mouse spleen was injected 30
10 minutes previously with *Bacillus anthracis*, *Bacillus thuringiensis*, *Bacillus subtilis* spores and labeled with blue, green and red antibodies, respectively.

DETAILED DESCRIPTION OF THE INVENTION

15

In the present invention, panels of monoclonal antibodies which are highly specific and can discriminate between spores of the *Bacillus* family including the strategically important *Bacillus anthracis* (anthrax) were isolated and characterized. The amino acid sequences
20 of these anti-spore antibodies were determined from the nucleotide sequences of the coding genes and smaller peptide molecules were derived from these antibodies which can also bind *Bacillus* spores.

The present invention is directed to a monoclonal antibody highly specific for *Bacillus* spores. Preferably, *Bacillus* is selected from the group consisting of *Bacillus anthracis*, *Bacillus thuringiensis*, *Bacillus subtilis* and other bacilli closely related to
5 *Bacillus anthracis*. Preferably, the antibody is IgG. The present invention is also directed to a peptide derived from the monoclonal antibody highly specific for *Bacillus* spores. Preferably, the peptide can also bind the *Bacillus* spores specifically.

Also disclosed in the present invention is a method of
10 preparing a monoclonal antibody highly specific for *Bacillus* spores, comprising the steps of immunizing an animal with the *Bacillus* spores and fusing local lymph nodes of the animal.

The present invention also is directed to a method of detecting anthrax in a field sample using a monoclonal antibody
15 highly specific for *Bacillus anthracis* spores by contacting the sample with the monoclonal antibody and measuring the amount of binding of the antibody to the sample compared to an appropriate control.

The present invention also is directed to a method of detecting lethal *Bacillus* spores in a field sample using the peptide
20 derived from the monoclonal antibody highly specific for *Bacillus* spores by contacting the sample with the peptide and measuring the amount of binding of the peptide to the sample compared to an

appropriate control.

The following terms have the definitions set below.

As used herein, "hybridoma" refers to a continuously growing antibody-secreting cell line derived from the fusion of a specific normal antibody-forming B cell from an immunized mouse
5 with an immortal myeloma cell line. Hybridomas secrete monoclonal antibodies described herein.

As used herein, "homogeneous staining" refers to the uniform staining at a similar intensity of all spores in a given sample.

10 As used herein, "heterogeneous staining" refers to the staining of one or more populations of spores in a given sample.

As used herein, "unfixed untreated spores" refers to spores that are in their native state in water and not treated with any kind of fixation reagent such as formalin or glutaraldehyde or
15 paraformaldehyde.

As used herein, "two-color flow cytometric analysis" refers to the identification of spore subpopulations or other particles by fluorescence activated flow cytometry using two independent fluorochrome labeled antibodies.

20 As used herein, "limiting dilution" refers to the distributing of hybridoma cells into tissue culture plates such that less than 30% of the wells contain a growing clone. Each well, according to

the Poisson distribution, should contain the progeny of only one cell.

The following examples are given for the purpose of illustrating various embodiments of the invention and are not meant to limit the present invention in any fashion.

5

EXAMPLE 1

Animals

Eight to twelve-week-old BALB/c mice were purchased from Charles River Laboratories (Raleigh, NC) or bred in our mouse facility. BALB/c mice were used for immunization and obtaining feeder cells for subcloning of hybridomas, phenotypic analysis and functional studies. Mice were housed in accordance with institutional policies for animal care and usage.

15

EXAMPLE 2

Bacterial Spores

B. subtilis spores were provided by Dr. Chuck Turnbough. *B. anthracis* spores were obtained from Dr. Joany J. Jackman at USAMIRID and *B. thuringiensis* spores were obtained from Abbott Laboratories.

EXAMPLE 3

Antibody Production: Immunization and Fusion

Six-week-old female BALB/c mice were inoculated with
5 5×10^8 spores emulsified in complete Freund's adjuvant at day 0, and
then repeatedly with spores in saline at days 3, 6, 9, 13, 17 and 20 in
subcutaneous sites in the rear legs and inguinal regions.

On day 21, popliteal, inguinal and iliac lymph nodes
draining sites of injection were removed, a lymphocyte suspension
10 was prepared and fused to P3x63Ag8.653 using a modification of the
method described by Kohler and Milstein. Fused cells were plated on
ten 96-well plates in DMEM supplemented with 20% fetal bovine
serum (FBS), 2 mmol/L L-glutamine, HAT medium, and FCS (from
HyClone Laboratories Inc., Logan, UT; other reagents from Sigma), and
15 placed in a 37°C incubator with 9% CO₂.

EXAMPLE 4

Primary Screening and Subcloning

20 Hybridoma supernatants were screened on spore
suspensions using two-color flow cytometric analysis. Binding of
secreted mouse Ig from supernatants to the spores was traced with

phycoerythrin (PE)-conjugated goat anti-mouse Ig (Southern Biotechnology Associates, Birmingham, AL). Data from stained cell samples were acquired using a FACScan or FACSCalibur flow cytometer with lysis II and Cell Quest software packages (Becton Dickinson, Mountain View, CA) and analyzed with WinList 2.01 (Verity Software House, Inc.) and WinMDI 2.0 software programs (Trotter@scripps.edu).

EXAMPLE 5

10

ELISA

Flat bottom ELISA plates (E.I.A.A/2 plates, Costar) were coated with poly-L-Lysine (50 µg) for 30 minutes and a suspension of spores at 2×10^8 /ml (40 µl) in distilled water were allowed to dry on the plates overnight. Supernatants were added and after incubation developed with goat anti-mouse Ig. Between each step, the plate was washed five times with PBS. The plate was developed with alkaline phosphatase substrate (Sigma, St. Louis, MO) (1 mg/ml) in substrate buffer (pH 9). For quantitative ELISA, mouse antibody of known concentration was used as a standard in each plate and OD₄₀₅ values of plates were read by a Titertek Multiskan Plus MKII spectrophotometer (Flow, McLean, VA). Antibody concentration was

determined using an ELISALITE program (Meddata, New York, NY).

EXAMPLE 6

5 Antibody Purification and Conjugation

Pure anti-spore antibodies were prepared from bulk hybridoma cultures by protein G chromatography. FITC and phycoerythrin conjugates were prepared using standard procedures.

10 EXAMPLE 7

Immunofluorescence and Immunohistochemical Analysis of Tissue Sections and Cytocentrifuge Preparations

Spleens embedded in OCT compound (Lab-Tek Products, Naperville, IL) were flash frozen in liquid nitrogen. Frozen sections were cut, air dried, fixed in ice-cold acetone, blocked with normal horse serum, and macrophages stained with MOMA-1 (rat, IgG2a, 10 µg/ml, from Dr. Georg Kraal), each developed with biotin-conjugated goat anti-rat IgG (SBA). Next, the sections were blocked with normal
15 rat serum followed by anti-spore reagents and secondary reagents and streptavidin AMCA (Vector Laboratories, Burlingame, CA). Spore
20 suspension in distilled water were dried onto poly-L-Lysine treated

glass slides for 2 hours at 37°C, blocked with 1% BSA and PBS and stained with antibodies for microscopy of spore suspensions.

Tissue sections and slides with dried spores were washed and mounted in Fluormount G (SBA, Birmingham, AL) and viewed
5 with a Leica/Leitz DMRB fluorescence microscope equipped with appropriate filter cubes (Chromatechnology, Battleboro, VT). Images were acquired with a C5810 series digital color camera (Hamamatsu Photonic System, Bridgewater, NJ) and processed with Adobe Photo Shop and IP LAB Spectrum software (Signal Analytics Software.
10 Vienna, VA).

EXAMPLE 8

DNA Sequencing Analysis

15 V_H and V_K gene sequencing was carried out from cDNA isolated from hybridomas. To make cDNA, total RNA was isolated from hybridomas using guanidinium thiocyanate-phenol-chloroform extraction. The cDNA was synthesized using an oligo-dT primer followed by PCR using a C_μ 3' primer (SEQ ID NO: 1) and V_H 7183-
20 specific primer (SEQ ID NO: 2) for the heavy chains or a C_k 3' primer (SEQ ID NO: 3) and a degenerate V_K 5' primer (SEQ ID NO: 4) for the light chains. The PCR amplified DNA was cloned into

Bluescript II KS and subjected to sequencing using a Sequenase Kit (Stratagene, La Jolla, CA). The DNA sequences were analyzed using the DNASTar program.

5

EXAMPLE 9

Antibodies to *B. subtilis*

Two immunization were made, one with fixed spores which gave only 3/192 (1.6%) monoclonal antibodies (mAbs) reactive with spores; and the other with unfixed spores which gave 95/384 (25%) (mAbs) reactive with spores, another 89 (20%) weakly reactive. These 576 clones were then tested against other spore components and 15 reacted with NAD synthetase, 6 with RNA polymerase, 5 with cot TC.2 with SSPC, and 1 with cse60 by ELISA. Totally, 136 clones were reactive with spores or purified/recombinant components.

Among the clones reactive with the intact spores, certain patterns were observed: (1) two clones appeared to dramatically alter the FSC/SSC profile of spores on flow cytometry; (2) some clones reacted with germinated, but not with ungerminated spores; and (3) the majority had homogenous staining of germinated, but heterogeneous staining of ungerminated spores (Figure 1).

96 clones of hybridomas reactive with *B. subtilis* were

picked up and grown on a new plate. They include two negative clones, the clones reactive with purified proteins or peptides and clones reactive with spores representing different patterns. These antibodies were tested on the vegetative forms of *B. subtilis* (i.e., live
5 bacteria) and were found to be negative (Figure 2). They also did not react with two other species of spore-forming *Bacilli* (Figure 3). Isotyping of the antibodies produced by these clones revealed that many (55/96) use λ light chains. Additionally, it was also unusual that 4 of these antibodies use α heavy chains.

10 All 96 clones were subcloned by limiting dilution and tested by flow cytometry. 68/96 were still reactive with spores and all except one were monoclonal. The reactive clones can be basically separated into two groups: those reactive with all spores and the other reactive with subsets of spores. Since these antibodies are of
15 different isotypes, multiple parametric flow cytometric analysis could be done next. These important results showed that fixation of spores did not permit production of antibodies to the intact native spores and it was only when unfixed untreated spores were used to immunize mice could many highly specific antibodies to *B. subtilis* be
20 isolated.

EXAMPLE 10

Sequence Analysis of Monoclonal Antibody to *B. subtilis*

The striking over usage of λ light chains in the antibodies
5 led to sequencing the heavy and light chains of the genes from
hybridomas synthesizing the *B. subtilis* spore specific antibodies to
obtain an idea of the heterogeneity of antibodies generated. The
heavy chains revealed remarkable homogeneity of V_H gene usage in
that all hybridomas used a member of V_H7183 (MOPC21) family. This
10 member $V_H7183.6$ appears to be the most unique member of this
family in the framework three (FR3) region as shown in Figure 4. The
CDR3 region was diverse in nearly all cases and used variable D_H and
 J_H genes. These results suggest that there is a very strong selection
for the use of this V_H gene despite the similarities inherent in the
15 family members of this family. λ light chain sequence showed the
exclusive use of $V\lambda1$ $C\lambda1$ with different CDR3 regions. Six of these
were selected for further study and characterization.

EXAMPLE 11

20

Isolation of FR3 Peptides Which Bind to Spores

Based on the sequences of V_H genes utilized in

antibodies against *Bacillus subtilis* spores, two peptides were designed: one corresponding to the consensus sequence of these antibodies in the framework 3 region (Peptide Anti-spore: SEQ ID NO: 5), and the other corresponding to the consensus sequences of the 5 7183 V_H gene family to which the particular V_H gene belongs (Peptide 7183 consensus: SEQ ID NO: 6).

The carboxyl-terminal cysteine was added for fluorochrome conjugation. Both peptides were conjugated with phycoerythrin, and tested for their ability to bind *Bacillus subtilis* 10 spores. 7183 consensus peptide was designed to be a control. It was found that the peptides derived from the anti-spore antibody stained brightly at 2 µg/ml (1 µM), while the consensus peptide stained spores at 200 µg/ml (100 µM) (Figure 5). Thus the peptide derived from the spore specific nucleotide derived antibody sequence bind 15 strongly and specifically to *B. subtilis*.

EXAMPLE 12

Serum Antibody Response to *B. subtilis* Spore Immunization

20 The immune response to *Bacillus subtilis* spores was characterized in mice. BALB/c mice were immunized with either spores or PBS (control). The mice were bled at 1, 2 and 3 weeks after

immunization. Serum antibodies of different isotypes specific for spores were quantitated using ELISA. It was found that (1) immune responses peaked at 1 week; (2) light chain-containing antibodies account for about 30% of total spore-specific immunoglobulins; and
5 (3) in contrast to all other isotypes, IgG3 antibodies continue to increase over the 3-week period. These findings confirmed the hybridoma analysis that the immune response to *B. subtilis* spores is dominated by a particular set of B cell clones.

10

EXAMPLE 13

Monoclonal Antibodies to *B. anthracis*

Mice were immunized with a 50:50 mix of heavily irradiated (4×10^6 Gy) *Bacillus anthracis* spores of the Ames and
15 Sterne strains, generated hybridomas, and screened for antibody production by FACS analysis. About 60 hybridomas were selected for further characterization. A similar pattern of reactive antibodies was obtained with some of these panels binding 100% of *B. anthracis* spores. As seen in Figure 6, a representative profile of more than 36
20 anti-anthrax antibodies which stain all spores but were not at all reactive with *B. subtilis* and *B. thuringiensis* spores. The V_H gene sequences were determined and are shown in Figure 7. Again a

similar conservation in V_H usage was found similar to what was found in antibodies to *B. subtilis*. In this case, one of the two V_H genes is from V_H7183 and the other from the V_HJ558 family predominates. A third V_H gene is from the V_HQ52 family.

5

EXAMPLE 14

Monoclonal Antibodies to *B. thuringiensis*

A similar strategy was used to isolate and characterize
10 ~100 antibody forming hybridomas which reacted with *B. thuringiensis*. Again the pattern was similar with all reacting with *B. thuringiensis* but not *B. subtilis* or *B. anthracis*. These antibodies were cloned and are sequenced. The discriminatory ability of antibodies is shown in Figures 8 and 9 where it is possible to clearly discriminate
15 three distinct spore staining by fluorescence in a mixture of the three kinds of spores *in vitro* and *in vivo*.

EXAMPLE 15

20 Discussion

The work presented here disclosed panels of antibodies which are highly specific and can discriminate between spores of the

Bacillus family including the strategically important *B. anthracis* (anthrax). This is the first time such antibodies have been isolated and characterized. The reagents used in the various Divisions of the Armed services for testing were not monoclonal. They were made in
5 sheep and other species against *B. subtilis* and *B. anthracis*.

The antibodies disclosed in the present invention are unique because of several reasons: (1) the spores were not fixed with glutaraldehyde or formalin (which chemically modifies the spores) before immunization; (2) these are monoclonal antibodies made by
10 immunizing and fusing local lymph nodes. Such a procedure has not been used in the past. The few monoclonal antibodies described before have been of the IgM isotypes which are more difficult antibodies to use and are more cross-reactive, i.e., react with spores other than *B. anthracis*, while the monoclonal antibodies disclosed
15 herein are IgG. IgM antibodies of this kind are useless in instruments designed to give positive results for anthrax spores in the field, since such antibodies will also detect harmless spores such as *B. subtilis* which is ubiquitous in the environment.

The amino acid sequences of these anti-spore antibodies
20 were also analyzed, which allows one to design and make smaller peptide molecules which can also bind spores. These will be more rugged molecules than the large antibody molecule and can be used in

other kinds of detectors. Such peptides are totally unique in their binding to *Bacillus* spores.

There are numerous government (services and intelligence), as well as private groups trying to make instruments that are small, portable and highly accurate in their detection of small numbers of potentially lethal spores such as anthrax. The monoclonal antibodies presented here could play a critical role in their instrument development program. Such findings will be significant in detecting air and water containing anthrax spores for civilian and military use.

Any patents or publications mentioned in this specification are indicative of the levels of those skilled in the art to which the invention pertains. These patents and publications are herein incorporated by reference to the same extent as if each individual publication was specifically and individually indicated to be incorporated by reference.

One skilled in the art will readily appreciate that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as well as those inherent therein. The present examples along with the methods, procedures, treatments, molecules, and specific compounds described herein are presently representative of preferred embodiments, are exemplary.

and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention as defined by the scope of the claims.

WHAT IS CLAIMED IS:

1. A monoclonal antibody highly specific for *Bacillus* spores.

5

2. The monoclonal antibody of claim 1, wherein said antibody is IgG.

10

3. The monoclonal antibody of claim 1, wherein said *Bacillus* is selected from the group consisting of *Bacillus anthracis*, *Bacillus thuringiensis*, *Bacillus subtilis*, and bacilli related to *Bacillus anthracis*.

15

4. A peptide derived from the monoclonal antibody of claim 1

5. The peptide of claim 4 binds *Bacillus* spores specifically.

20

6. A method of preparing a monoclonal antibody highly

specific for *Bacillus* spores, comprising the steps of:

immunizing an animal with said *Bacillus* spores; and
fusing local lymph nodes of said animal.

5 7. The method of claim 6, wherein said monoclonal
antibody is IgG.

8. The method of claim 6, wherein said *Bacillus* is
selected from the group consisting of *Bacillus anthracis*, *Bacillus*
10 *thuringiensis*, *Bacillus subtilis*, and bacilli related to *Bacillus anthracis*.

9. The method of claim 6, wherein said animal is a
mouse.

15 10. A method of detecting *Bacillus* spores in a field
sample using a monoclonal antibody specific for said spores,
comprising the step of:

contacting said sample with the monoclonal antibody of
claim 1.

20 11. The method of claim 10, wherein said monoclonal
antibody is IgG.

12. The method of claim 10, wherein said *Bacillus* is selected from the group consisting of *Bacillus anthracis*, *Bacillus thuringiensis*, *Bacillus subtilis*, and bacilli related to *Bacillus anthracis*.

5 13. A method of detecting *Bacillus* spores in a field sample, comprising the step of:

 contacting said sample with said peptide of claim 4.

 14. The method of claim 13, wherein said peptide binds
10 *Bacillus* spores specifically.

Antibodies with different reactivities for spores

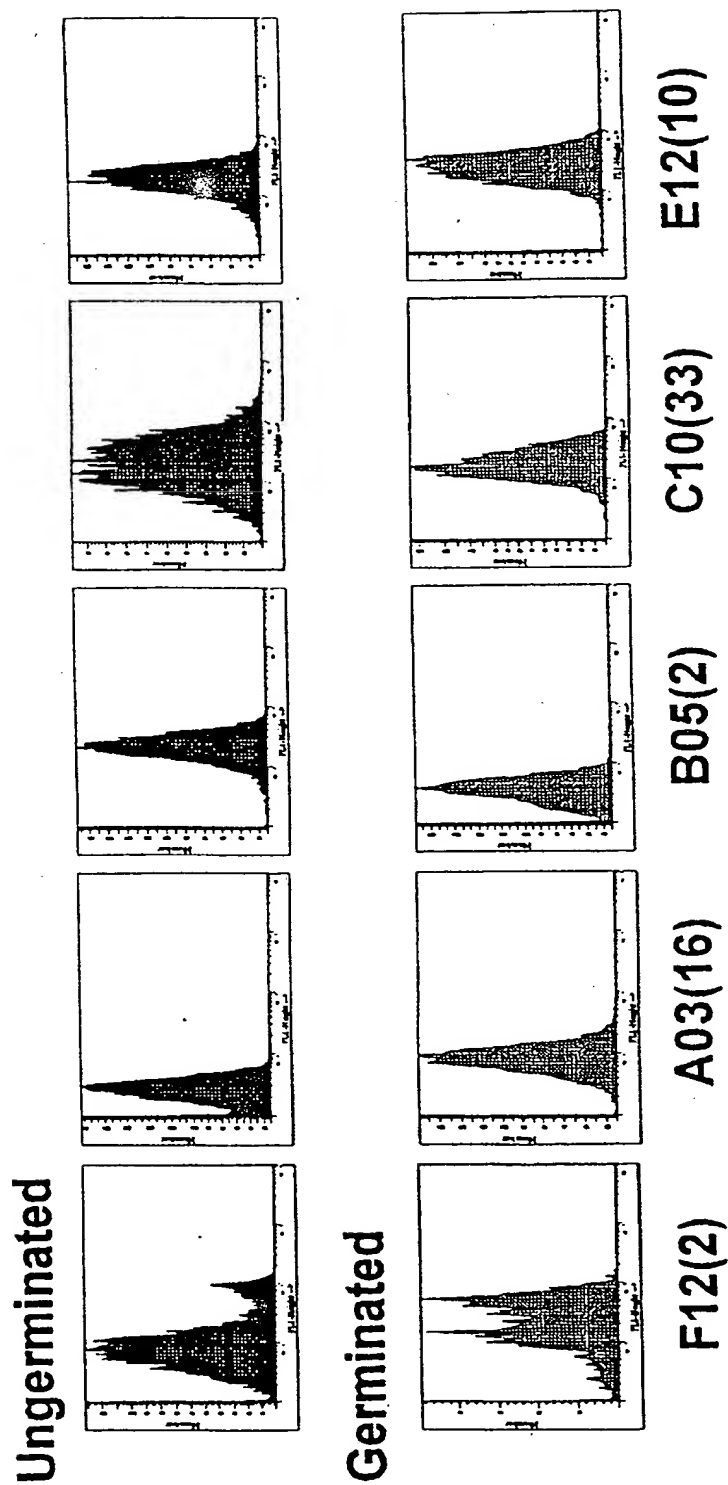
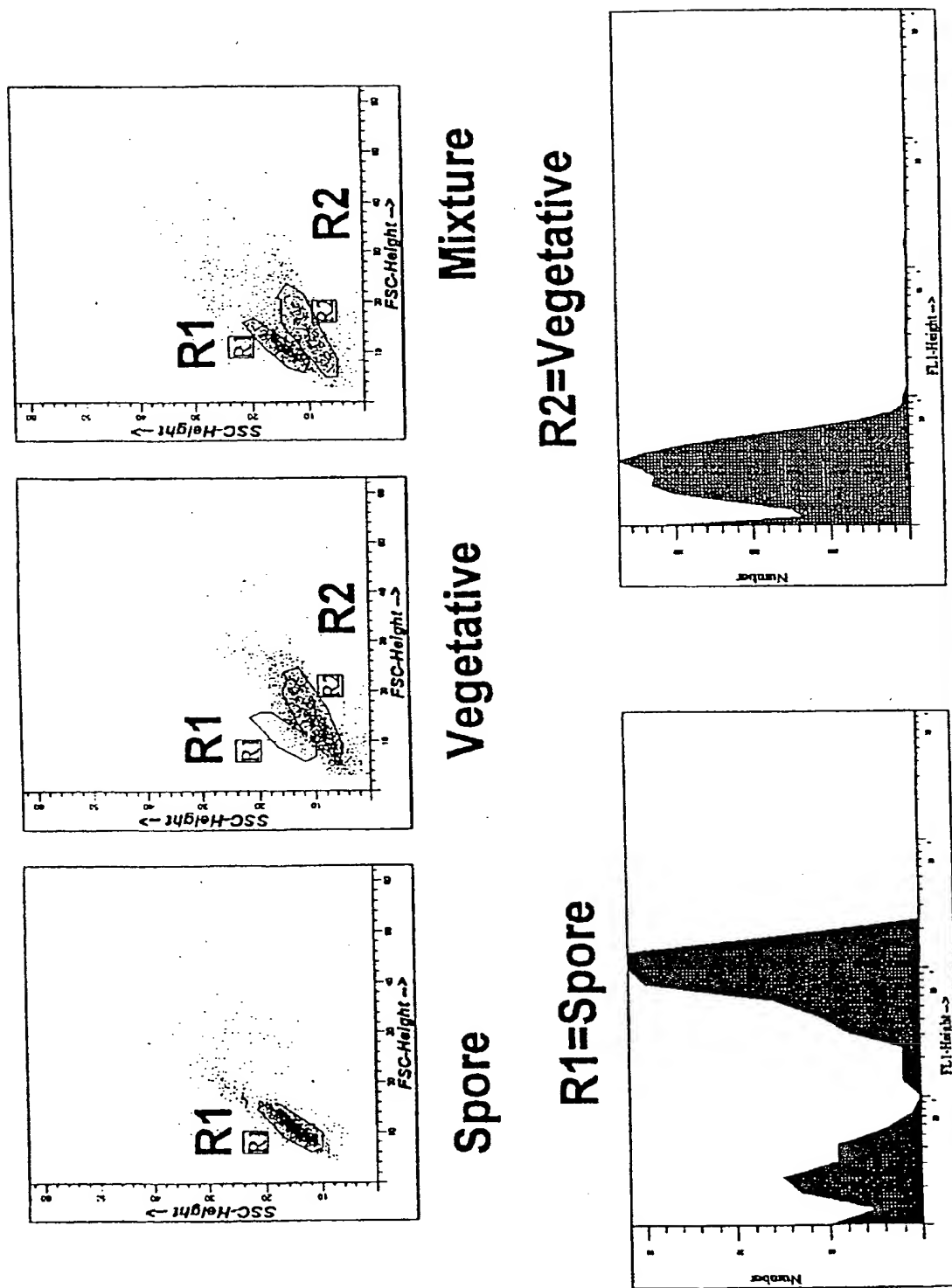


FIG. 1

Anti-spore mAbs do not react with vegetative bacteria

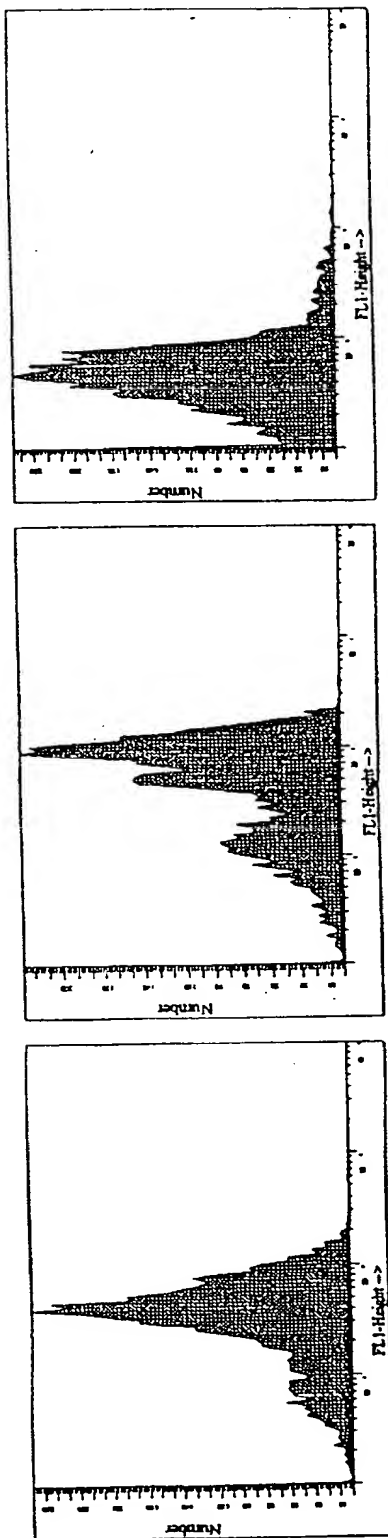


D12

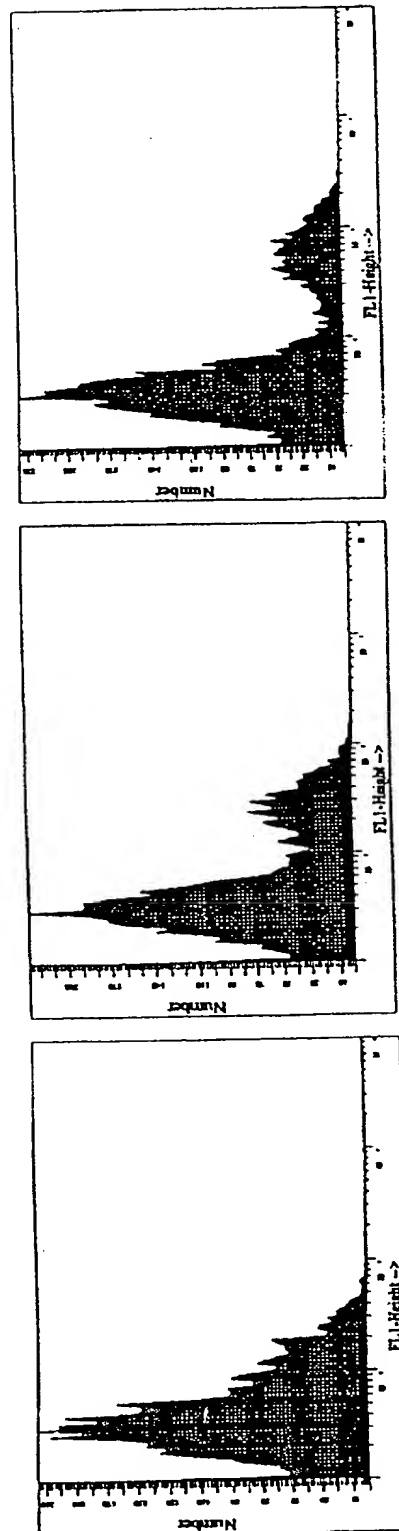
FIG. 2

Most antibodies react specifically with spores of *Bacillus subtilis*

B. subtilis



B. thuringiensis



C11 like most

G6

F11 and other two

FIG. 3

g07	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
g04	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
g06	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
d06	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
a07	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
e11	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
a05	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
e07	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
d12	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
d04	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
f10	1	GGLVQPGGSRKLS	CAASGFTFSSFGMHVVRQAP	KEKGLEWVAYISSGSSTIYYADTVKGRF
g07	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...RGTGTRYEDYNGQGT
g04	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...YR...MDYNGQGT
g06	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...GHDYGYSGRYEDVWGAGT
d06	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...GHYGYSGRYEDVWGAGT
a07	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...WEVTRYEDVWGAGT
11	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...TVVRRAMDYNGQGT
a05	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...GSKRAIDYNGQGT
e07	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...DLRTEFYNGQGT
d12	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...LLRAMDYNGQGT
d04	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...HYGTNYVRAMDYNGQGT
f10	61	TISRDNPKN	TLFLQMTSLRSEDTAMYCAR	...HYDEGPHWYEDVWGAGT

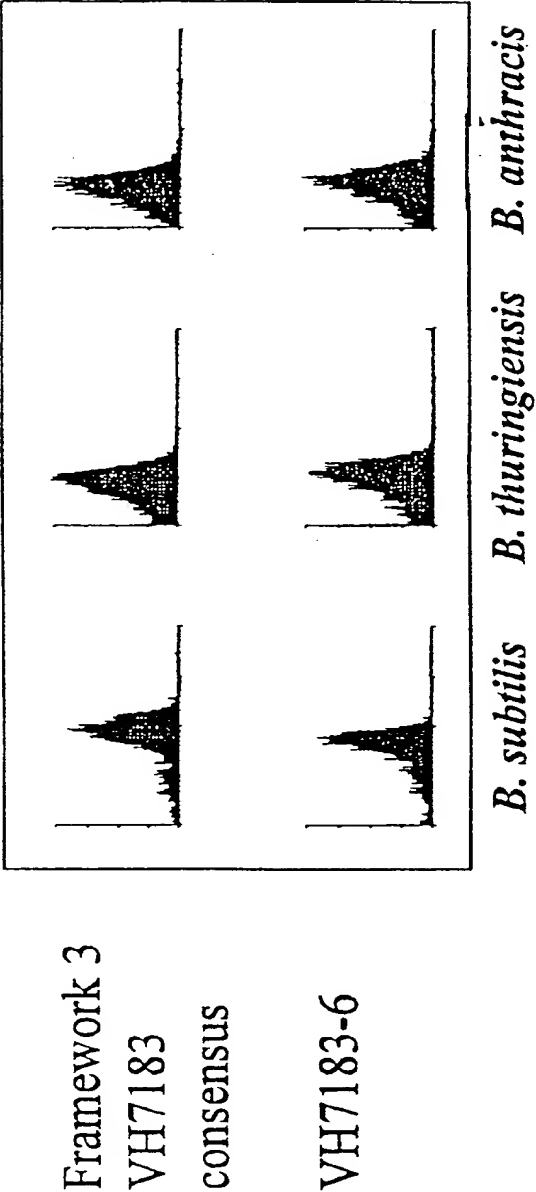
FIG. 4-1

g07	117	PNVPLVSCESPLSDKNLVGCLDPD~
g04	115	PNVPLVSCESPLSDKNLVGCLDPD~
g06	121	SNVPLVSCESPLSDKNLVGCLDPD~
d06	121	PSVYPLVPGCIDTSGSS.VTLGCLVKAT
a07	117	PSVYPLVPGCSDTSGSS.VTLGCLVKAT
e11	119	PSVYPLVPGCIDTSGSS.VTLGCLVKAT
a05	118	PSVYPLVPGCSDTSGSS.VTLGCLVKAT
e07	116	PSVYPLAPVCGDTGSS.VTLGCLVKGY
d12	115	PSVYPLAPVCGDTGSS.VTLGCLVKGY
d04	121	PSVYPLAPGCGDTGSS.VTLGCLVNGY
f10	121	PSVYPLAPGCGDTGSS.VTLGCLVKGY

g07	SEQ ID NO: 7	a05	SEQ ID NO: 13
g04	SEQ ID NO: 8	e07	SEQ ID NO: 14
g06	SEQ ID NO: 9	d12	SEQ ID NO: 15
d06	SEQ ID NO: 10	d04	SEQ ID NO: 16
a07	SEQ ID NO: 11	f10	SEQ ID NO: 17
e11	SEQ ID NO: 12		

FIG. 4-2

Ab-derived Peptides Specifically Bind
Bacillus subtilis Spores



VH7183-6	RFTISRDNPKNTLFLQMT	SEQIDNO: 5
VH7183 consensus	RFTISRDNKNTLLYLQMS	SEQIDNO: 6

FIG. 5

Anti-Bacillus anthracis Ab Specifically Bind
Bacillus anthracis Spores

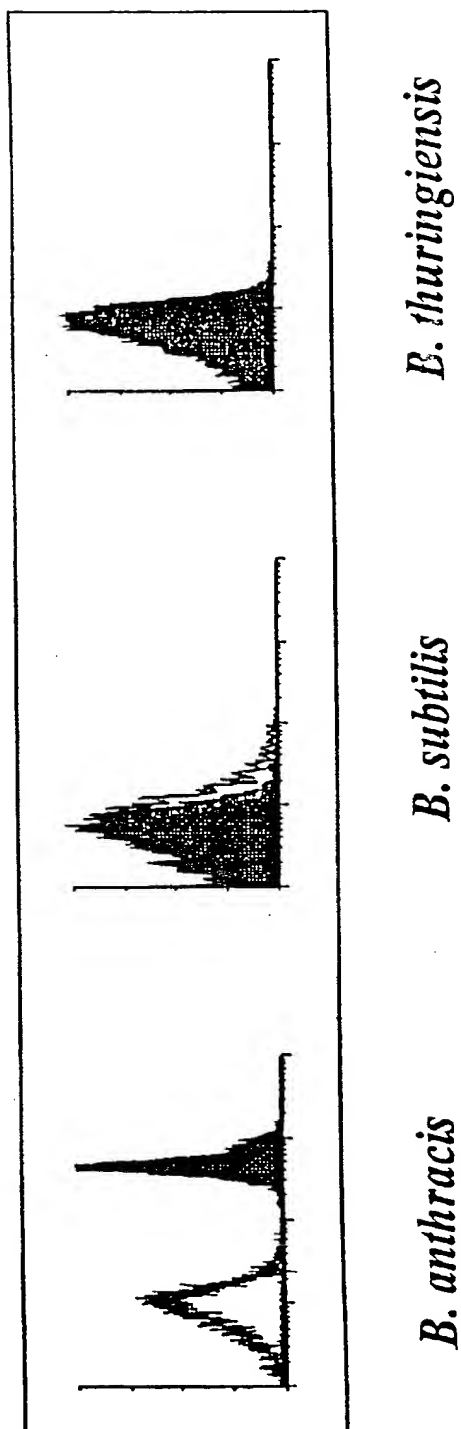


FIG. 6

[illegible]

FIG. 7-1

2	SEQIDNO: 18	10-2	SEQIDNO: 25
3	SEQIDNO: 19	22-1	SEQIDNO: 26
4	SEQIDNO: 20	13-3	SEQIDNO: 27
9-1	SEQIDNO: 21	8-3	SEQIDNO: 28
7-1	SEQIDNO: 22	6-1	SEQIDNO: 29
24-2	SEQIDNO: 23	3-1	SEQIDNO: 30
21-4	SEQIDNO: 24	1	SEQIDNO: 31

FIG. 7-2

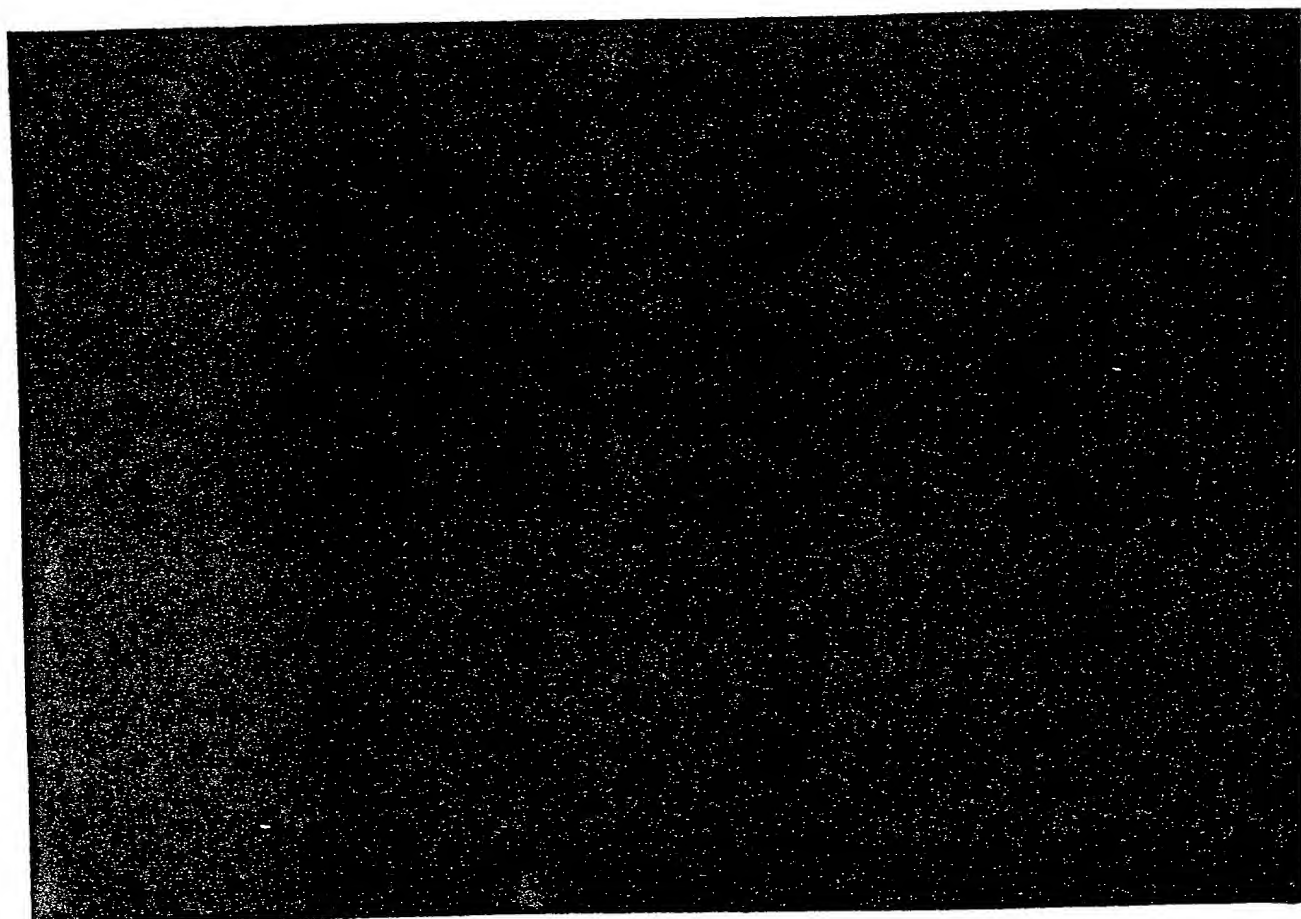


FIG. 8

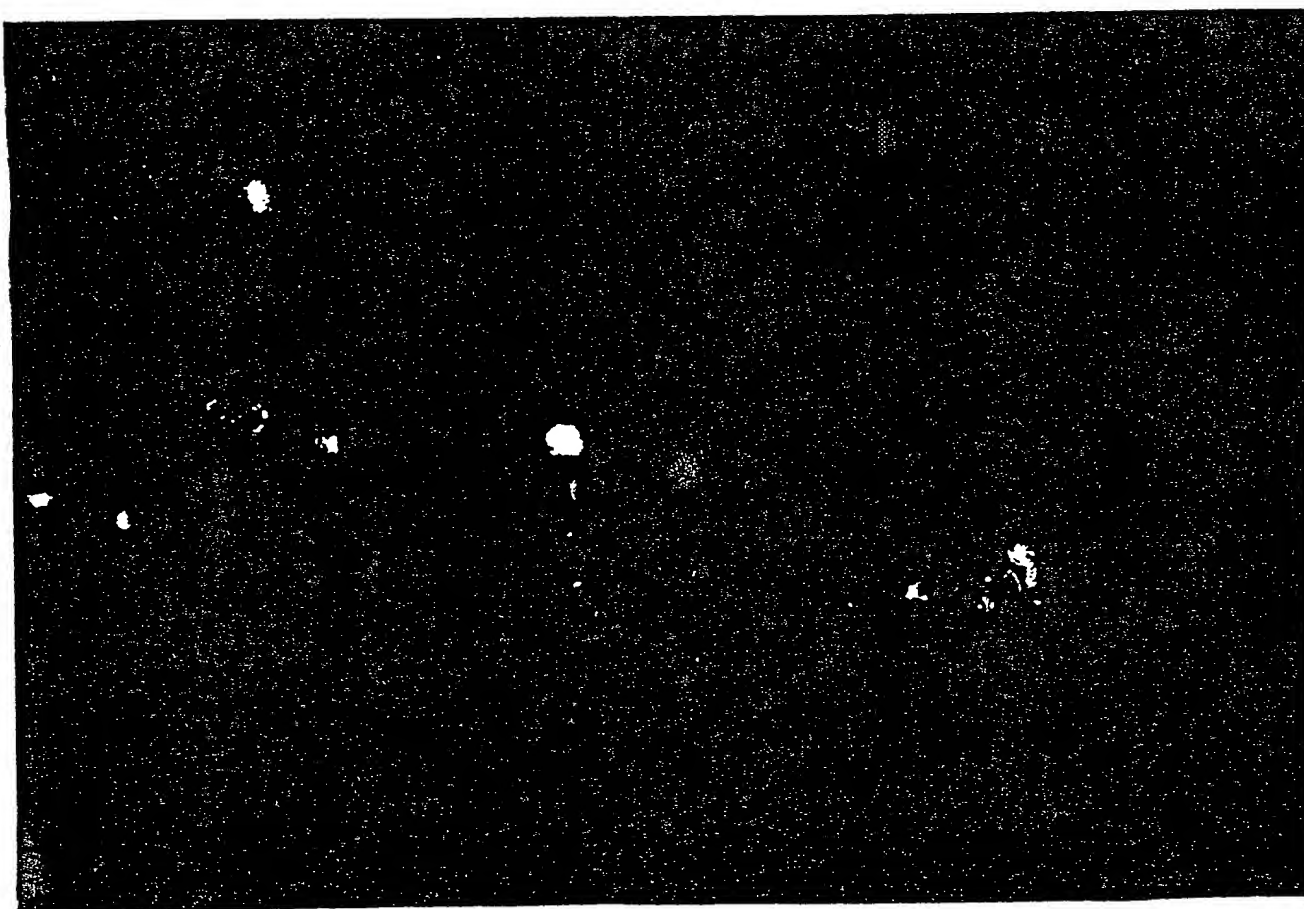


FIG. 9

SEQUENCE LISTING

<110> Kearney, John F.
 <120> Monoclonal Antibodies Specific for Anthrax
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 <140>
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 <150> US 09/069,628
 <151> 1998-04-29
 <160> 31

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 <223> C κ 3' primer used to amplify V_K cDNA
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<210> 4
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 <212> DNA
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 <223> Ck 5' primer used to amplify V_K cDNA
 <400> 4

gccatgggtpr tqlwlmtsac ccagtctcca 30

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 <223> Peptide Anti-spore VH7183-6 amino acid sequence
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 region of *B. subtilis* antibodies.
 <400> 5

Arg Phe Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu

5 10 15

Gln Met Thr

<210> 6
 <211> 18
 <212> PRT
 <213> artificial sequence
 <220>
 <223> Peptide 7183 consensus sequence corresponding to
 the consensus sequences of the 7183 VH gene family.
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Arg Phe Thr Ile Ser Arg Asp Asn Ala Lys Asn Thr Leu Tyr Leu

5 10 15

Gln Met Ser

<210> 7
 <211> 143

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 <223> Hybridoma g07 amino acid sequence of framework
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Gly	Gly	Leu	Val	Gln	Pro	Gly	Gly	Ser	Arg	Lys	Leu	Ser	Cys	Ala
				5						10				15
Ala	Ser	Gly	Phe	Thr	Phe	Ser	Ser	Phe	Gly	Met	His	Trp	Val	Arg
				20						25				30
Gln	Ala	Pro	Glu	Lys	Gly	Leu	Glu	Trp	Val	Ala	Tyr	Ile	Ser	Ser
				35						40				45
Gly	Ser	Ser	Thr	Ile	Tyr	Tyr	Ala	Asp	Thr	Val	Lys	Gly	Arg	Phe
				50						55				60
Thr	Ile	Ser	Arg	Asp	Asn	Pro	Lys	Asn	Thr	Leu	Phe	Leu	Gln	Met
				65						70				75
Thr	Ser	Leu	Arg	Ser	Glu	Asp	Thr	Val	Met	Tyr	Tyr	Cys	Ala	Arg
				80						85				90
Arg	Gly	Thr	Gly	Thr	Arg	Tyr	Phe	Asp	Tyr	Trp	Gly	Gln	Gly	Ala
				95						100				105
Thr	Leu	Thr	Val	Ser	Ser	Glu	Ser	Gln	Ser	Phe	Pro	Asn	Val	Phe
				110						115				120
Pro	Leu	Val	Ser	Cys	Glu	Ser	Pro	Leu	Ser	Asp	Lys	Asn	Leu	Val
				125						130				135
Ala	Met	Gly	Cys	Leu	Asp	Pro	Asp							
				140										

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<210>	9
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three (FR3) region of heavy chain gene.

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Gly Gly Leu Val Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala		
5	10	15
Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg		
20	25	30
Leu Ala Pro Asp Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Ser		
35	40	45
Gly Ser Ser Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe		
50	55	60
Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met		
65	70	75
Thr Ser Leu Arg Ser Asp Asp Thr Ala Met Tyr Tyr Cys Ala Arg		
80	85	90
Ser Gly His Asp Tyr Gly Tyr Ser Arg Gly Tyr Phe Asp Val Trp		
95	100	105
Gly Ala Gly Thr Thr Val Thr Val Ser Ser Glu Ser Gln Ser Phe		
110	115	120
Ser Asn Val Phe Pro Leu Val Ser Cys Glu Ser Pro Leu Ser Asp		
125	130	135
Lys Asn Leu Val Ala Met Gly Cys Leu Asp Pro Asp		
140	145	

<210> 10

<211> 147

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<220>

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three (FR3) region of heavy chain gene.

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           5              10              15
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           20              25              30
Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Ser
           35              40              45
Gly Ser Ser Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe
           50              55              60
Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met
           65              70              75
Thr Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg
           80              85              90
Ser Gly His Tyr Tyr Gly Tyr Ser Arg Gly Tyr Phe Asp Val Trp
           95              100             105
Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ala Thr Thr Thr Ala
           110             115             120
Pro Ser Val Tyr Pro Leu Val Pro Gly Cys Ile Asp Thr Ser Gly
           125             130             135
Ser Ser Val Thr Leu Gly Cys Leu Val Lys Ala Thr
           140             145

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Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg		
	20	25 30
Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Ser		
	35	40 45
Gly Ser Ser Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe		
	50	55 60
Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met		
	65	70 75
Thr Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg		
	80	85 90
Tyr Asp Thr Thr Val Val Ala Arg Ala Met Asp Tyr Trp Gly Gln		
	95	100 105
Gly Thr Ser Val Thr Val Ser Ser Ala Thr Thr Thr Ala Pro Ser		
	110	115 120
Val Tyr Pro Leu Val Pro Gly Cys Ile Asp Thr Ser Gly Ser Ser		
	125	130 135
Val Thr Leu Gly Cys Leu Val Lys Ala Thr		
	140	145

<210> 13

<211> 144

<212> PRT

<213> artificial sequence

<220>

<223> Hybridoma a05 amino acid sequence of framework
three (FR3) region of heavy chain gene.

<400> 13

Gly Gly Leu Val Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala		
	5	10 15

Ala Ser Gly Phe Thr Phe Ser Thr Phe Gly Val His Trp Val Arg		
	20	25 30

<210>	14
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<223>	Hybridoma e07 amino acid sequence of framework three (FR3) region of heavy chain gene.
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			5						10					15
Ala	Ser	Gly	Phe	Thr	Phe	Ser	Thr	Phe	Gly	Met	His	Trp	Val	Arg
			20						25					30
Gln	Ala	Pro	Glu	Lys	Gly	Leu	Glu	Trp	Val	Ala	Tyr	Ile	Ser	Ser
			35						40					45

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Gly Ser Ser Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe
 50 55 60
 Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met
 65 70 75
 Thr Asn Leu Thr Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg
 80 85 90
 Trp Asp Ala Leu Arg Thr Phe Ala Tyr Trp Gly Gln Gly Thr Leu
 95 100 105
 Val Thr Val Ser Ala Ala Lys Thr Thr Ala Pro Ser Val Tyr Pro
 110 115 120
 Leu Ala Pro Val Cys Gly Asp Thr Thr Gly Ser Ser Val Thr Leu
 125 130 135
 Gly Cys Leu Val Lys Gly Tyr
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 <223> Hybridoma d12 amino acid sequence of framework
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 <400> 15

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 5 10 15
 Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg
 20 25 30
 Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Ser
 35 40 45
 Gly Ser Ser Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe
 50 55 60

Thr	Ile	Ser	Arg	Asp	Asn	Pro	Lys	Asn	Thr	Leu	Phe	Leu	Gln	Met
				65					70					75
Thr	Ser	Leu	Thr	Ser	Glu	Asp	Thr	Ala	Met	Tyr	Tyr	Cys	Ala	Arg
				80					85					90
Trp	Leu	Leu	Arg	Ala	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Ser	Val
				95					100					105
Thr	Val	Ser	Ser	Ala	Lys	Thr	Thr	Ala	Pro	Ser	Val	Tyr	Pro	Leu
				110					115					120
Ala	Pro	Val	Cys	Gly	Asp	Thr	Thr	Gly	Ser	Ser	Val	Thr	Leu	Arg
				125					130					135
Cys	Leu	Val	Lys	Gly	Tyr									
				140										

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 <400> 16

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				5					10					15
Ala	Ser	Gly	Phe	Thr	Phe	Ser	Thr	Phe	Gly	Met	His	Trp	Val	Arg
				20					25					30
Gln	Ala	Pro	Glu	Lys	Gly	Leu	Glu	Trp	Val	Ala	Tyr	Ile	Ser	Ser
				35					40					45
Gly	Ser	Ser	Thr	Ile	Tyr	Tyr	Ala	Asp	Thr	Val	Lys	Gly	Arg	Phe
				50					55					60
Thr	Ile	Ser	Arg	Asp	Asn	Pro	Lys	Asn	Thr	Leu	Phe	Leu	Gln	Met
				65					70					75

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Thr Ser Leu Thr Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg
 80 85 90
 Trp His Tyr Tyr Gly Thr Asn Tyr Val Arg Ala Met Asp Tyr Trp
 95 100 105
 Gly Gln Gly Thr Ser Val Thr Val Ser Ser Ala Lys Thr Thr Ala
 110 115 120
 Pro Ser Val Tyr Pro Leu Ala Pro Gly Cys Gly Asp Thr Thr Gly
 125 130 135
 Ser Ser Leu Thr Leu Gly Cys Leu Val Asn Gly Tyr
 140 145

<210> 17

<211> 147

<212> PRT

<213> artificial sequence

<220>

<223> Hybridoma f10 amino acid sequence of framework
 three (FR3) region of heavy chain gene.

<400> 17

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 5 10 15
 Ala Ser Gly Phe Thr Phe Ser Thr Phe Gly Met His Trp Val Arg
 20 25 30
 Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Ser
 35 40 45
 Gly Ser Ser Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe
 50 55 60
 Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met
 65 70 75
 Thr Ser Leu Thr Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg
 80 85 90

Ser Ser Val Thr Leu Gly Cys Leu Val Lys Gly Tyr
140 145

<210>	18
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<212>	PRT
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<223>	<i>B. anthracis</i> monoclonal antibody 2 VH gene sequence
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Thr Tyr Pro Ile Pro Ile Arg

5

<210>	19
<211>	7
<212>	PRT
<213>	artificial sequence
<220>	
<223>	<i>B. anthracis</i> monoclonal antibody 3 VH gene sequence
<400>	19

Thr Tyr Pro Ile Pro Phe Arg

$\langle 210 \rangle$	20
$\langle 211 \rangle$	7

<212> PRT
 <213> artificial sequence
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 <223> *B. anthracis* monoclonal antibody 4 VH gene
 sequence
 <400> 20

Thr Tyr Pro Val Pro His Arg

5

<210> 21
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 <212> PRT
 <213> artificial sequence
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 sequence
 <400> 21

Pro	Ser	Gln	Ser	Leu	Ser	Ile	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser
				5					10					15
Leu	Ser	Gly	Tyr	Ser	Val	His	Trp	Val	Arg	Gln	Arg	Pro	Gly	Lys
				20					25					30
Gly	Leu	Glu	Cys	Leu	Gly	Met	Ile	Trp	Gly	Val	Gly	Ser	Thr	Asp
				35					40					45
Tyr	Asn	Ser	Ala	Leu	Lys	Ser	Arg	Leu	Ser	Ile	Ser	Lys	Asp	Asn
				50					55					60
Ser	Lys	Ser	Gln	Val	Phe	Leu	Lys	Met	Asn	Ser	Leu	Gln	Thr	Asp
				65					70					75
Asp	Thr	Ala	Met	Tyr	Tyr	Cys	Ala	Arg	Asp	Tyr	Tyr	Gly	Asn	Tyr
				80					85					90
Val	Trp	Tyr	Phe	Asp	Val	Trp	Gly	Ala	Gly	Thr	Thr	Val	Thr	Val
				95					100					105

<210>	22
<211>	129
<212>	PRT
<213>	artificial sequence
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<223>	<i>B. anthracis</i> monoclonal antibody 7-1 VH gene sequence
<400>	22

SEQ 15/23

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125

<210> 23

<211> 143

<212> PRT

<213> artificial sequence

<220>

<223> *B. anthracis* monoclonal antibody 24-2 VH gene
sequence

<400> 23

Gly Gly Leu Val Lys Pro Gly Gly Ser Leu Lys Leu Ser Cys Ala

5 10 15

Ala Ser Gly Phe Thr Phe Ser Ser Tyr Ala Met Ser Trp Val Arg

20 25 30

Gln Thr Pro Glu Lys Arg Leu Glu Trp Val Ala Thr Ile Ser Ser

35 40 45

Gly Gly Ser Tyr Thr Tyr Tyr Pro Asp Ser Val Lys Gly Arg Phe

50 55 60

Thr Ile Ser Arg Asp Asn Ala Lys Asn Thr Leu Tyr Leu Gln Met

65 70 75

Ser Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg

80 85 90

Gln Gly Leu Arg Arg Tyr Ala Met Asp Tyr Trp Gly Gln Gly Thr

95 100 105

Ser Val Thr Val Ser Ser Ala Lys Thr Thr Pro Pro Ser Val Tyr

110 115 120

Pro Leu Ala Pro Gly Phe Gly Asp Thr Thr Gly Ser Ser Val Thr

125 130 135

Leu Gly Cys Leu Val Lys Gly Tyr

140

SEQ 16/23

<210> 24
 <211> 142
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 sequence
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 5 10 15
 Ala Ser Gly Phe Thr Phe Ser Ser Tyr Ala Met Ser Trp Val Arg
 20 25 30
 Gln Thr Pro Glu Lys Arg Leu Glu Trp Val Ala Thr Ile Ser Ser
 35 40 45
 Gly Gly Ser Tyr Thr Tyr Tyr Pro Asp Ser Val Lys Gly Arg Phe
 50 55 60
 Thr Ile Ser Ile Asp Asn Ala Lys Asn Thr Leu Tyr Leu Gln Met
 65 70 75
 Ser Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg
 80 85 90
 Gln Gly Leu Arg Arg Val Ala Met Asp Tyr Trp Gly Gln Gly Thr
 95 100 105
 Ser Val Thr Val Ser Ser Ala Lys Thr Thr Ala Pro Ser Val Tyr
 110 115 120
 Gln Leu Ala Pro Gly Phe Gly Asp Thr Thr Gly Ser Ser Val Thr
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 sequence
 <400> 25

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				5					10					15
Ala	Ser	Gly	Phe	Thr	Phe	Ser	Ser	Tyr	Ala	Met	Pro	Trp	Val	Arg
				20					25					30
Gln	Thr	Pro	Glu	Lys	Arg	Leu	Glu	Trp	Val	Ala	Thr	Ile	Arg	Ser
				35					40					45
Gly	Gly	Ser	Tyr	Thr	Tyr	Tyr	Pro	Asp	Ser	Val	Lys	Gly	Arg	Phe
				50					55					60
Thr	Ile	Ser	Arg	Asp	Asn	Ala	Lys	Asn	Thr	Leu	Tyr	Leu	Gln	Met
				65					70					75
Ser	Ser	Leu	Arg	Ser	Glu	Asp	Thr	Ala	Met	Tyr	Tyr	Cys	Pro	Ile
				80					85					90
Tyr	Asp	Gly	His	Ala	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Ser	Val
				95					100					105
Thr	Val	Ser	Ser	Ala	Thr	Thr	Thr	Ala	Pro	Ser	Val	Tyr	Pro	Leu
				110					115					120
Val	Pro	Gly	Cys	Ala	Asp	Thr	Thr	Gly	Ser	Ser	Val	Thr	Leu	Gly
				125					130					135
Cys	Leu	Val	Lys	Gly	Tyr									
				140										

<210> 26
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 <212> PRT
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<220>

<223> *B. anthracis* monoclonal antibody 22-1 VH gene
sequence

<400> 26

Gly	Asp	Leu	Val	Lys	Pro	Gly	Gly	Ser	Leu	Lys	Leu	Ser	Cys	Ala			
				5					10					15			
Ala	Ser	Gly	Phe	Thr	Phe	Ser	Ser	Tyr	Gly	Met	Ser	Trp	Val	Arg			
				20					25					30			
Gln	Thr	Pro	Asp	Lys	Arg	Leu	Glu	Trp	Val	Ala	Thr	Ile	Ser	Ser			
				35					40					45			
Gly	Gly	Ser	Tyr	Thr	Tyr	Tyr	Pro	Asp	Ser	Val	Lys	Gly	Arg	Phe			
				50					55					60			
Thr	Ile	Ser	Arg	Asp	Asn	Ala	Lys	Asn	Thr	Leu	Tyr	Leu	Gln	Met			
				65					70					75			
Ser	Ser	Leu	Lys	Ser	Glu	Asp	Thr	Ala	Met	Tyr	Tyr	Cys	Ala	Arg			
				80					85					90			
Arg	Gly	Ile	Thr	Thr	Ala	Ile	Tyr	Ala	Met	Asp	Tyr	Trp	Gly	Gln			
				95					100					105			
Gly	Thr	Ser	Val	Thr	Val	Ser	Ser	Glu	Ser	Gln	Ser	Phe	Pro	Asn			
				110					115					120			
Val	Phe	Pro	Leu	Val	Ser	Cys	Glu	Ser	Pro	Leu	Ser	Asp	Lys	Asn			
				125					130					135			
Leu	Val	Ala	Met	Gly	Cys	Leu	Ala	Arg	Asp								
				140					145								

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SEQ 19/23

<223> *B. anthracis* monoclonal antibody 13-3 VH gene
sequence

<400> 27

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				5					10					15
Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Arg	Tyr	Thr	Met	His	Trp	Val	Lys
				20					25					30
Gln	Arg	Pro	Gly	Gln	Gly	Leu	Glu	Trp	Ile	Gly	Tyr	Ile	Asn	Pro
				35					40					45
Ser	Ser	Gly	Tyr	Thr	Asn	Tyr	Asn	Gln	Lys	Phe	Lys	Asp	Lys	Ala
				50					55					60
Thr	Leu	Thr	Ala	Asp	Lys	Ser	Ser	Arg	Thr	Ala	Tyr	Met	Gln	Leu
				65					70					75
Ser	Ser	Leu	Thr	Ser	Glu	Asp	Ser	Ala	Val	Tyr	Tyr	Cys	Ala	Arg
				80					85					90
Val	Thr	Ala	Arg	Tyr	Ala	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Ser
				95					100					105
Val	Thr	Val	Ser	Ser	Ala	Lys	Thr	Thr	Ala	Pro	Ser	Val	Tyr	Pro
				110					115					120
Leu	Ala	Pro	Val	Cys	Gly	Asp	Thr	Thr	Gly	Ser	Ser	Val	Thr	Leu
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 Lys Gln Arg Pro Gly His Gly Leu Glu Trp Ile Gly Asp Ile Tyr
 35 40 45
 Pro Gly Gly Gly Tyr Thr Asn Tyr Asn Glu Lys Phe Lys Gly Lys
 50 55 60
 Ala Thr Leu Thr Ala Asp Thr Ser Ser Ser Thr Ala Tyr Met Gln
 65 70 75
 Leu Ser Ser Leu Thr Ser Glu Asp Ser Ala Ile Tyr Tyr Cys Ala
 80 85 90
 Arg Gly Asn Leu Gly Asp Tyr Trp Gly Gln Gly Thr Thr Leu Thr
 95 100 105
 Val Ser Ser Ala Lys Thr Thr Ala Pro Ser Val Tyr Pro Leu Ala
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 125 130 135
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Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Ser	Tyr	Trp	Met	His	Trp	Val
			20						25					30
Lys	Gln	Arg	Pro	Gly	Gln	Gly	Leu	Glu	Trp	Ile	Gly	Asn	Ile	Asn
			35						40					45
Pro	Ser	Pro	Gly	Tyr	Thr	Glu	Tyr	Asn	Gln	Lys	Phe	Lys	Asp	Lys
			50						55					60
Gly	Thr	Leu	Thr	Ala	Asp	Lys	Ser	Ser	Ser	Thr	Ala	Tyr	Met	Gln
			65						70					75
Leu	Ser	Ser	Leu	Thr	Ser	Glu	Asp	Ser	Ala	Val	Tyr	Tyr	Cys	Ala
			80						85					90
Arg	Ile	Gly	Ser	Gly	Tyr	Val	Gly	Tyr	Ala	Met	Asp	Tyr	Trp	Gly
			95						100					105
Gln	Gly	Thr	Ser	Val	Thr	Val	Ser	Ser	Glu	Ser	Gln	Ser	Phe	Pro
			110						115					120
Asn	Val	Phe	Pro	Leu	Val	Ser	Cys	Glu	Ser	Pro	Leu	Ser	Glu	Lys
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sequence

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Thr Ser Gln Asn Val Arg Thr

5

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/09122

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C12N 5/06; A61K 35/00, 39/102 37/00

US CL : 424/ 93.462, 200.1, 290; 435/ 69.7, 340; 514/2; 800/205

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : Please See Extra Sheet.

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
MANUAL OF MICONIOLOGY AND BIOTECHNOLOGY, SPORES, SPORE ANTIGENS, INFECTION AND IMMUNOLOGY, MICROB. PATHOG.

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5,254,799 A (DE GREVE et al.) 19 October 1993, col. 15-18.	1-5
Y,P	US 5,753,222 A (MARRONE et al.) 19 May 1998, col. 1 lines 37-49.	1-5
A	US 5,686,113 A (SPEAKER et al.) 11 November 1997, col. 4-8, col. 12-13, col. 17 lines 36-48.	1-5
Y	WALKER et al. Immunology of Sores and Sporeforms, Anitgens of the Spore and Location of Spore and Vegatative Anitgens With Labeled Anitobdies. 1972, Vol. 5, pages 321-337, see entire document.	1-5 and 10- 14



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
B earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*g* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

18 JULY 1999

Date of mailing of the international search report

02 SEP 1999

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/09122

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	LITTLE et al. Comparative Efficacy of Bacillus-Anthraxis Live Spore Vaccine and Protective Antigen vaccine Against Anthrax in the Guinea-Pig. Infect. and Immun. May 1986, Vol. 53, No. 2, pages 509-512.	1-14

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/09122

B. FIELDS SEARCHED

Minimum documentation searched

Classification System: U.S.

424/ 93.462, 200.1, 290

435/ 69.7, 340

514/ 2

800/205